Clinical Cancer Research



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Clin Cancer Res 2011;17:6029-6039. Published OnlineFirst August 1, 2011.

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Imaging, Diagnosis, Prognosis

Prognostic Impact of Δ TAp73 Isoform Levels and Their Target Genes in Colon Cancer Patients

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Abstract

Purpose: Cumulative data support the role of Δ TAp73 variants in tumorigenic processes such as drug resistance. We evaluate the impact of *TP73* isoforms and their putative target genes *ABCB1*, *HMGB1*, and *CASP1* on the survival of colon cancer patients and the correlation between their expressions.

Experimental Design: We determined in 77 colon cancer patients the expression of $\Delta Ex2p73$, $\Delta Ex2/3p73$, $\Delta Np73$, TAp73, ABCB1, HMGB1, and CASP1 by quantitative real-time reverse transcriptase-PCR. Tumor characteristics, disease-free survival, and overall survival (OS) were examined in each patient. Functional experiments were carried out to check whether ectopic expression of $\Delta Np73$ modifies the proliferation, drug resistance, migration, and invasion properties of colon tumor cells and the expression of ABCB1, HMGB1, and CASP1.

Results: Positive correlations were observed between the expression levels of $\Delta TAp73$ variants and *HMGB1*. Furthermore, a trend was observed for *ABCB1*. Overexpression of $\Delta Ex2/3p73$ and $\Delta Np73$ isoforms was significantly associated with advanced stages (P = 0.04 and P = 0.03, respectively) and predicted shortened OS (P = 0.04 and P = 0.05, respectively). High levels of *ABCB1* and *HMGB1* were associated with shorter OS (P = 0.04 and P = 0.05, respectively). Multivariate analysis showed that, in addition to the tumor stage, *ABCB1* and *HMGB1* had independent relationships with OS (P = 0.008). Ectopic expression of $\Delta Np73$ was associated with an increase in proliferation and drug resistance.

Conclusions: The positive correlation between $\Delta TAp73$ variants and *HMGB1* and *ABCB1* expression supports them as TP73 targets. The fact that upregulation of $\Delta TAp73$ isoforms was associated with shortened OS, increase in proliferation, and drug resistance confirms their oncogenic role and plausible value as prognostic markers. *ABCB1* and *HMGB1*, putative $\Delta TAp73$ target genes, strongly predict OS in an independent manner, making clear the importance of studying downstream TP73 targets that could predict the outcome of colon cancer patients better than $\Delta TAp73$ variants themselves do. *Clin Cancer Res;* 17(18); 6029–39. ©2011 AACR.

Introduction

TP73, a member of the *p53* family, is expressed in multiple variants. TAp73 isoforms (full-length) have tumor-suppressor potential (1), whereas Δ TAp73 variants

doi: 10.1158/1078-0432.CCR-10-2388

(ΔEx2p73, ΔEx2/3p73, ΔNp73, and ΔNCp73), lacking the transactivation domain, show oncogenic properties (2-7). TP53 and TP73 share significant structural and functional homology (8), although some evidence shows that their roles differ in human tumorigenesis. The 2 genes are activated through different pathways after DNA damage, and are capable of inducing cell-cycle arrest and cell death. Unlike TP53, inactivating mutations of TP73 are extremely rare in human tumors (9). Moreover, although TP73 can activate some TP53-responsive genes to varying degrees, such as those induced after DNA damage (10, 11), recent analyses showed that p73 has its own set of target genes (11, 12), indicating unique and overlapping functions for this family. Further complexity is revealed by the fact that the members of the TP53 family can transactivate common target genes but through the recognition of distinct binding elements (11). In addition, several reports have indicated that $\Delta Np73$ acts downstream from TP53 and TAp73 as a transcriptional negative regulator that

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Translational Relevance

The tumor-suppressor and/or oncogenic functions of TP73 isoforms have been intensely debated recently. The publication of an article in 2010 on the specific role of $\Delta TAp73$ in knockout mice strongly supports their role as oncogenes in the tumorigenic process. Thus, there is currently increasing interest in unraveling the mechanisms that underlie the oncogenic potential of $\Delta TAp73$ isoforms. Our report is the first that shows the prognostic value of $\Delta TAp73$ variants and their target genes involved in drug resistance in colon cancer patients. The positive correlation found in our colon cancer series between ΔTAp73 variants and HMGB1 and ABCB1 expression supports them as TP73 targets in vivo. The upregulation of these 2 genes after ectopic expression of $\Delta Np73$ in colon tumor cells also sustains this statement. The fact that $\Delta TAp73$ isoforms are associated with shortened overall survival (OS) and with increase in proliferation and drug resistance confirms their oncogenic role and plausible value as prognostic markers. ABCB1 and HMGB1 strongly predict OS in an independent manner, making clear the importance of studying downstream TP73 targets that could predict the outcome of colon cancer patients better than Δ TAp73 themselves do.

competes for binding motifs in target promoters (4, 13). The latest finding is supported by the evidence that loss of Δ Np73 makes some target promoters more accessible to TP53 and TAp73 (14).

TP53 knockout (KO) mice develop normally, but show high predisposition to the development of spontaneous tumors (15). In contrast, *TP73*-null mice (all *TP73* isoforms absent) have developmental neurologic and immunologic defects with early deaths, but no tumor predisposition (16). Specifically, mice lacking *TAp73* show neurologic defects and develop spontaneous malignancies (17), whereas $\Delta TAp73$ -null mice display signs of neurodegeneration and impaired tumorigenic capability (14).

Several studies have reported the expression status of TP73 isoforms in human tumors (18-22), but only a few showed the involvement of $\Delta TAp73$ isoforms in the prognosis of patients (23-31). The failure or lack of tumor response is mainly due to drug resistance mechanisms. In clinical oncology, multidrug resistance remains an unresolved problem. TP53 and TAp73 isoforms have functions in the apoptotic response to drug-induced DNA damage (32-34). $\Delta Np73$ acts as a dominant-negative inhibitor of TAp73 and wild-type p53 (4), inhibiting drug-induced apoptosis (25, 35). In addition, TP73 regulates the expression of genes associated with drug resistance. Specifically, upregulation of the activity of ABCB1, HMGB1, and CASP1 promoters by p73 variants has been described previously (35–38). MDR1 is a P-glycoprotein that acts as an energydependent drug-efflux pump and is often overexpressed in multidrug-resistant tumor cells (39). HMGB1 is involved in several biologic processes, including invasiveness,

transcription, DNA repair, and drug resistance mechanisms (40, 41). Lastly, CASP-1 plays an important role in several apoptosis pathways (38) and has been described as being downregulated in several tumor types (42, 43). In addition, overexpression of *ABCB1* and *HMGB1* has been associated with poor prognosis of cancer patients (44–46).

This is the first report that shows the prognostic value of $\Delta TAp73$ variants and their target genes involved in drug resistance in patients diagnosed with colon cancer. The positive correlation found between $\Delta TAp73$ variants and *HMGB1* and *ABCB1* expression supports them as TP73 targets. The fact that $\Delta TAp73$ isoforms are associated with shortened overall survival (OS) confirms their oncogenic role and plausible value as prognostic markers. *ABCB1* and *HMGB1* strongly predict OS in an independent manner, making clear the importance of studying downstream TP73 targets that could predict the outcome of colon cancer patients better than $\Delta TAp73$ themselves do.

Materials and Methods

Patients, tumor samples, and nucleic acid extraction

The present study, approved by the research ethics board of the Puerta de Hierro Majadahonda University Hospital (Madrid, Spain), was based on a consecutive series of 77 patients undergoing surgery for colorectal cancer between January 2001 and January 2003. All colon cancer patients were considered sporadic cases because no clinical antecedents of familial adenomatous polyposis (FAP) were reported and those with clinical criteria of hereditary nonpolyposis colorectal cancer (Amsterdam criteria) were excluded. Both normal and tumor tissue samples were obtained sequentially, immediately after surgery, snapfrozen in liquid nitrogen, and stored at -80° C until further processing.

All tumors were histologically examined by a pathologist to confirm the diagnosis of colon cancer, verify the presence of tumor, select those samples with at least 75% tumor tissue, and establish the pathologic stage.

RNA was extracted from approximately 30 mg of colon tumor and normal tissue samples using the RNeasy Mini Kit (Qiagen Inc.). After extraction, RNA was quantified spectrophotometrically.

Real-time PCR

mRNA levels were detected in the normal and tumor counterpart samples by a relative quantification approach in which the amount of the targets is expressed in relation to the geometric average of the 3 reference housekeeping genes, as described in detail elsewhere (20). The relative concentrations of the target and the reference genes were calculated by interpolation, using a standard curve of each gene plotted from a serial dilution of a cDNA prepared from the RNA of an individual expressing the specific analyzed gene. The expression level of the target gene in a patient was calculated as a ratio: target in tumor tissue to target in normal tissue (T:N). For the synthesis of the firststrand cDNA, 400 ng of RNA was reverse-transcribed, using the Gold RNA PCR Core Kit (Applied Biosystems) according to the manufacturer's instructions. Random hexamers were used as primers for cDNA synthesis.

Real-time PCR (quantitative PCR) was carried out in a Light Cycler apparatus (Roche Diagnostics) using the Light-Cycler-FastStart DNA Master SYBR Green I Kit (Roche Diagnostics). Each reaction was carried out in a final volume of 20 μ L containing 2 μ L of the cDNA product sample, 0.5 μ mol/L of each primer, and 1× reaction mix including FastStar DNA polymerase, reaction buffer, deoxyribonucleotide triphosphates, and SYBR green.

Thermal cycling for all genes was initiated with a denaturing step at 95°C for 10 minutes and then subjected to 40 cycles of PCR (denaturing at 94°C for 10 seconds, annealing at a different temperature for each gene—67°C for 5 seconds for *ABCB1*, 58°C for 5 seconds for *HMGB1*, and 62°C for 4 seconds for *CASP1*—and elongation at 72°C for 5 seconds, in which fluorescence was acquired). At the end of the PCR cycles, melting curve analyses were conducted, followed by sequencing, to validate the generation of the specific PCR product expected.

Primer sets for $\Delta Ex2p73$, $\Delta Ex2/3p73$, $\Delta Np73$, and TAp73and the conditions for each reaction have been described elsewhere (20). Primer pairs for *ABCB1*, *HMGB1*, and *CASP1* were designed using Primer Express version 2.0 (Applied Biosystems). The following primers were used: forward, 5'CTATGCATCTTATGCTCTGGCC3'; reverse, 5'CCTGTCCAACACTAAAAGCCC3' for *ABCB1*; forward, 5'ACCCAGATGCTTCAGTCAACTTC3'; reverse, 5'TGCCATATCTTCAAATTTTCCTTTC3' for *HMGB1*; and forward, 5'AGTTACCTGGCAGGGACGCT3'; reverse, 5'TGGAAAGGAAGAAAGTACTCCTTGA 3' for *CASP1*.

Proliferation, migration, invasion, and drug resistance experiments

The colon cancer cells HCT116 were obtained from the American Type Culture Collection and maintained in Dulbecco's Modified Eagle Medium (DMEM; Lonza Group Ltd). Cells were seeded in triplicate and transiently transfected with a pcDNA plasmid encoding Δ Np73 or the empty vector (kindly provided by Dr. Marín, Instituto de Biomedicina, Universidad de León, Spain), using Lipofectamine 2000 (Invitrogen) according to the manufacturer's instructions. At 24, 48, 72, and 96 hours post-transfection, different fractions were kept to preserve cells and isolate RNA and/or protein. RNA samples were submitted to a DNAase treatment for evaluation of the levels of Δ Np73, HMGB1, ABCB1, and CASP1.

Proliferation was evaluated by 3 different approaches: First, cell density was assessed with a cell-counter apparatus (Digital Bio). Second, 2×10^4 colon cells were seeded in quadruplicates in 96-well E-plates to carry out an MTT cell proliferation assay (Cayman Chemical Company). At 24, 48, and 72 hours posttransfection, MTT reagent was added and absorbance was measured on a microplate reader at 570 nm (Multiskan Ex; Thermo Scientific). The RT-CES microelectronic cell sensor system (ACEA) was used for analysis. Cells were placed on the reader in the incubator for continuous recording of impedance (every 10 minutes for 96 hours) as reflected by cell index (47). Cells were transfected when attached (15 hours after seeding), and impedance changes are shown 12 hours after transfection (after 27 hours of the beginning of the process).

For drug resistance experiments, HCT116 cells were treated with 100 μ mol/L oxaliplatin for 36 hours. Subsequently, floating and adherent cells were trypsinized and checked for viability by flow cytometry using the Annexin V-FTTC Apoptosis Detection Kit (BD Pharmingen). Specifically, cells were resuspended in 1× binding buffer at a concentration of 1 × 10⁶ cells/mL. Two hundred microliters of the cell suspension was transferred to a 5-mL polypropylene tube, and 5 μ L each of propidium iodide (PI; 50 μ g/mL stock) and Annexin V-fluorescein isothiocyanate (FTTC) was added simultaneously. Cells were mixed and incubated at room temperature for 15 minutes in the dark. Cells were analyzed within 30 minutes.

HCT116 cells were cultured on $8.0-\mu$ m pore Transwells (Corning Inc.) previously covered with either 0.5% gelatin for migration assays or with Matrigel matrix (125 µg/ mL; BD Biosciences) for invasion assays. Before culture, cells were labeled with Cell Tracker Green (CMFDA C2925; Invitrogen). After several time points, from 8 to 72 hours, cells adhering to the lower surface of the filter were recovered by trypsinization and counted by fluorescence with the Wallac Plate Reader (Ex: 485 nm; Em: 535 nm; Perkin Elmer Life Science) by interpolation using a standard curve.

TP53 analysis

TP53 immunophenotypic analysis in the colon tissue samples was conducted according to standard procedures, with overnight incubation in the presence of the cl1801 mouse monoclonal antibody (Oncogene Sciences). Immunodetection was carried out with peroxidase-labeled streptavidin biotin (LSA; DAKO) using diaminobenzidine chromogen as substrate. All immunostaining was done using the TechMate 500 (DAKO) automatic immunostaining device. The cl1801 mouse monoclonal antibody was used because of its ability to detect up to 89% of *TP53* point mutations (48). Tissue samples exhibiting definitive nuclear (or nuclear and cytoplasmic) staining in more than 10% of the epithelial cells were considered positive for TP53. Cases displaying no nuclear staining were considered negative.

Clinicopathologic parameters

The following parameters were obtained from the medical records of the 77 patients: age, tumor size, tumor location, lymph node metastases, pathologic stage, histologic grade, and vascular invasion (VI). Pathologic stage was assessed by the tumor-node-metastasis (TNM) classification. Presence of lymph node metastases was evaluated by optical microscopy. No other immunohistochemical or molecular techniques were used. No patient received chemotherapeutic treatment before undergoing surgery.

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Patient follow-up

Clinical follow-up after diagnosis and surgery was based on periodic visits (every 3 months during the first year, every 6 months during the second year, and then yearly until relapse, in our medical oncology department, complemented by other periodic controls in health centers of our hospital), clinical and biochemical tests, and computed tomography scans. In addition, an ultrasonic study was done when liver function was impaired. OS and disease-free survival (DFS) were the study endpoints. OS was defined as the period from time of diagnosis until death. DFS was defined as the interval between diagnosis and first recurrence.

Statistical analysis

As the values of gene expression (T:N ratio) displayed nonnormal distribution (Kolmogorov–Smirnov test, Lillie-fors' correction), the data were normalized by log_{10} transformation. For the same reason, we used the geometric, rather than the arithmetic, average of the T:N ratio to describe the gene expression data.

Expression of *TP73* isoforms, *ABCB1*, *CASP1*, and *HMGB1* was divided into bicentiles and tertiles. The DFS analysis did not include the patients with pathologic stage IV disease. OS distribution was estimated by the Kaplan–Meier method (49), and differences between groups were tested using the log-rank test (50). Cox proportional hazard univariate and multivariate analyses were also conducted, including relative risk and 95% confidence intervals (CI). Finally, the Cox proportional risk regression model was fitted to data to estimate the independent prognostic importance of OS and DFS and confuser variables were analyzed (51). The basic assumptions of the model were evaluated (proportional hazards).

For statistical study of quantitative variables in the proliferation assays, the mean and SD were calculated. Student *t*-test was conducted to compare mean values of mock and Δ Np73 cells.

All *P* values were 2-sided, and values less than 0.05 were considered to indicate statistical significance. Analyses were conducted using the Statistical Package for Social Sciences version 14 (SPSS v.14).

Results

Association between TP73 isoform levels and tumor stage

Pathologic stage is the prognostic factor that has most clearly shown practical use in colorectal cancer. In a previous report of a series of 113 colorectal cancer patients, we found an association between tumor stage and expression levels of $\Delta Ex2/3p73$ and $\Delta Np73$ isoforms (20). Our current series of 77 patients is included in the aforementioned report, and we posited whether this association was maintained. $\Delta Ex2/3p73$ expression was significantly higher in stage IV (P = 0.04), with geometric averages of 0.24 for stage I, 0.25 for stage II, 0.16 for stage III, and 7 for stage IV. $\Delta Np73$ levels increased in parallel with stage (P = 0.03). The geometric average expressions were 0.009, 0.27, 0.36, and 5.33 in stages I, II, III, and IV, respectively.

Correlation between expression of TP73 variants and prognosis

The follow-up period of our series was the interval between surgery and the time of last medical appointment or death. As of October 2009, the series had been followed for a median of 70 months (range of follow-up, 3–104 months). During this period, 19 recurrences (24.3%) were recorded and 21 patients (27%) died. Description of the number of recurrences and deaths in the different categories for each variable is shown in Table 1.

Disease-free survival

The Kaplan–Meier and univariate analyses were conducted to determine the influence of stage and *TP73* isoform levels on DFS. No statistical associations were observed between *TP73* variant levels and DFS. As expected, tumor stage correlated in both statistical approaches with DFS (P = 0.002 and P = 0.02 for Kaplan–Meier and univariate analyses, respectively). Patients at stage III had a 5-year DFS rate of 59.6% (95% CI, 36.3–82.9); patients at stage II, 76.4% (95% CI, 57.6–95.2); and those at stage I, 100%. In the multivariate analysis, the pathologic stage was seen as a statistically supported factor in DFS prediction (P = 0.015).

Overall survival

In the final analysis, the 5-year OS for patients was 57% (95% CI, 43.5–70.5).

The tumor stage correlated in the Kaplan-Meier and univariate analyses with OS (P < 0.0001 and P < 0.0001, respectively). Patients at stage IV had a 5-year OS rate of 20% (95% CI, 0-55.1); patients at stage III, 32.4% (95% CI, 10.3-54.5); patients at stage II, 76.7% (95% CI, 62.4-91); and those at stage I, 87.7% (95% CI, 64.6-100). The Kaplan-Meier survival analysis revealed an association between OS and $\Delta Ex2/3p73$ expression when its levels were divided into bicentiles (P = 0.038; Fig. 1A). Patients with low $\Delta Ex2/3p73$ expression had a 5-year OS rate of 66.8% (95% CI, 47.2-86.4), whereas patients with high levels had a rate of 48.2% (95% CI, 31.1-65.3; Fig. 1B). A trend was observed in OS for the expression of $\Delta Np73$ (P = 0.06). Patients with low expression had a 5-year OS rate of 72.4% (95% CI, 56.9-87.9), whereas patients with high levels had a rate of 39.6% (95% CI, 17.3-61.9).

Correlation between TP73 isoform expression and mRNA levels of drug resistance related genes

Direct correlations were found between the levels of $\Delta Ex2p73$, $\Delta Ex2/3p73$, and $\Delta Np73$ and HMGB1 expression (Table 2). Similarly, a significant statistical trend was observed between $\Delta Ex2p73$, $\Delta Ex2/3p73$, and $\Delta Np73$ expression and ABCB1 levels (Table 2). No other correlations were identified.

Correlation between levels of drug resistance related genes and prognosis

Disease-free survival. Kaplan-Meier and univariate analyses were conducted to determine the influence of

| Variable | Category | Recurrences ($n = 19/77$) | % | Deaths (<i>n</i> = 21/77) | % |
|---|----------|-----------------------------|------|----------------------------|------|
| Stage | I | 0/9 | 0 | 1/9 | 11 |
| - | Ш | 7/42 | 16.6 | 5/42 | 12 |
| | III | 9/20 | 45 | 9/20 | 45 |
| | IV | 3/6 | 50 | 6/6 | 100 |
| Vascular invasion | No | 7/48 | 14.6 | 7/48 | 14.6 |
| | Yes | 12/29 | 41.4 | 14/29 | 48.3 |
| Tumor differentiation | Well | 8/50 | 16 | 11/50 | 22 |
| | Moderate | 10/22 | 45 | 9/22 | 41 |
| | Poor | 1/5 | 20 | 1/5 | 20 |
| LNM | No | 8/53 | 15 | 7/53 | 13 |
| | Yes | 11/24 | 46 | 14/24 | 58 |
| Bicentiles ∆ <i>Ex2p73</i> expression | Low | 10/38 | 26 | 9/38 | 24 |
| | High | 9/39 | 23 | 12/39 | 31 |
| Bicentiles ∆ <i>Ex2/3p73</i> expression | Low | 11/38 | 29 | 8/38 | 21 |
| | High | 8/39 | 21 | 13/39 | 33 |
| Bicentiles <i>ΔNp73</i> expression | Low | 11/38 | 29 | 8/38 | 21 |
| | High | 8/39 | 21 | 13/39 | 33 |
| Bicentiles TAp73 expression | Low | 11/38 | 29 | 12/38 | 32 |
| | High | 8/39 | 21 | 9/39 | 23 |
| Bicentiles ABCB1 expression | Low | 11/51 | 22 | 9/51 | 18 |
| | High | 8/26 | 31 | 12/26 | 46 |
| Bicentiles HMGB1 expression | Low | 3/26 | 11 | 4/26 | 15 |
| | High | 16/51 | 31 | 17/51 | 33 |
| Bicentiles CASP1 expression | Low | 10/38 | 26 | 12/38 | 32 |
| | High | 9/39 | 23 | 9/39 | 23 |

ABCB1, CASP1, and HMGB1 levels on DFS. No statistical associations were observed.

Overall survival. Patients were divided into bicentiles based on *ABCB1*, *CASP1*, and *HMGB1* levels. Because no differences between low and high levels were observed for

OS, we decided to divide patients into tertiles. Thus, patients presented low, median, or high levels of expression. No association was observed for the expression of *ABCB1* (P = 0.1). Patients with low expression had a 5-year OS rate of 74.9% (95% CI, 55.7–94.1); patients with

Figure 1. Influence of colon carcinoma levels of $\Delta Ex2/3p73$ (A) and $\Delta Np73$ (B) variants on OS, Kaplan–Meier curves, and *P* values. Expression was distributed in low and high levels by the median.



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| ∆ Ex2n73 | TAn73 |
|---|---------------|
| human colon cancer patients | |
| Table 2. Correlations between expression levels of p73 isoforms and HMGB1, ABCB1, a | and CASP1 for |

| | ∆ Ex2p73 | ∆Ex2/3p/3 | ∆ Np73 | TAp73 | |
|---|-----------------------------------|-----------------------------------|----------------------------------|-------|--|
| HMGB1 | <i>P</i> > 0.0001; <i>r</i> = 0.4 | <i>P</i> = 0.012; <i>r</i> = 0.28 | <i>P</i> = 0.04; <i>r</i> = 0.23 | NS | |
| ABCB1 | <i>P</i> = 0.06; <i>r</i> = 0.28 | <i>P</i> = 0.08; <i>r</i> = 0.25 | P = 0.08; r = 0.25 | NS | |
| CASP1 | NS | NS | NS | NS | |
| NOTE: Die selevieted hu englusie of veriences nie the Deersen exefficient | | | | | |

NOTE: *P* is calculated by analysis of variance; *r* is the Pearson coefficient. Abbreviation: NS, no statistically significant correlation.

median levels, a rate of 69.6% (95% CI, 50.8–88.4); and those with the highest levels, a rate of 35.5% (95% CI, 12.6–58.4; Fig. 2A). The Kaplan–Meier graph revealed similar behavior of median- and low-level tertiles (Fig. 2A). Thus, these patients were grouped as described above, and *ABCB1* expression was analyzed further with only 2 categories: low and high expression levels of *ABCB1*. When OS was analyzed in these 2 groups, a significant difference was observed, because patients with low *ABCB1* expression had a 5-year OS rate of 71.8% (95% CI, 58.3–85.3) and patients with high expression, a rate of 35.5% (95% CI, 12.6–58.4; P = 0.03; Fig. 2B).

No correlation was observed, either, for *HMGB1* expression (P = 0.1). Patients with low expression had a 5-year OS rate of 69.7% (95% CI, 44.2–95.2); patients with median levels, a rate of 45.2% (95% CI, 24–66.4); and those with the highest levels, a rate of 58.3% (95% CI, 38.5–78.1; Fig. 3A). The Kaplan–Meier graph revealed similar behavior of median- and high-level tertiles (Fig. 3A). Thus, these patients were grouped as above, and *HMGB1* expression was analyzed further with only 2 categories: low and high expression levels of *HMGB1*. When OS was analyzed in these 2 groups, a significant difference was observed, because patients with low *HMGB1* expression showed a 5-year OS rate of 69.7% (95% CI,

44.2–95.2) and patients with high expression, a rate of 51% (95% CI, 36.1–65.9; P = 0.04; Fig. 3B).

Similar results were found in the univariate analysis, in which expression levels of *ABCB1* and *HMGB1* were seen as a statistically supported factor in OS prediction (P = 0.04 and P = 0.05, respectively).

Multivariate analysis showed that tumor stage, *ABCB1*, and *HMGB1* had independent relationships with OS. When *TP73* isoform expression data were included in the multivariate analysis, tumor stage, *HMGB1*, and *ABCB1* levels again showed independent relationships with OS (Table 3).

Correlation between expression of TP73 variants and prognosis depending on TP53 status

Positive TP53 immunostaining (nuclear), suggesting *TP53* mutations, was observed in 53 out of 77 colon patients (70%)

Disease-free survival. Kaplan–Meier and univariate analyses were conducted to determine the influence of *TP73* isoforms on DFS depending on TP53 status. Patients were divided into bicentiles based on $\Delta Ex2p73$, $\Delta E2/3p73$, $\Delta Np73$, and *TAp73* levels. In those cases showing a positive immunostaining for TP53 (suggestive of mutation) a significant difference was observed with regard to $\Delta E2/3p73$



Figure 2. Influence of colon carcinoma levels of *ABCB1* on OS, Kaplan–Meier curves, and *P* values. A, *ABCB1* expression in colon cancer patients distributed by tertiles in low, median, and high levels. B, *ABCB1* expression in colon cancer patients distributed in 2 groups: low (new variable combining low and median levels) and high expression. Figure 3. Influence of colon carcinoma levels of *HMGB1* on OS, Kaplan–Meier curves, and *P* values. A, *HMGB1* expression in colon cancer patients distributed by tertiles in low, median, and high levels. B, *HMGB1* expression in colon cancer patients distributed in 2 groups: high (new variable combining median and high levels) and low expression.



because patients with low $\Delta E2/3p73$ expression had a 5year DFS rate of 71.2% (95% CI, 51.6–90.8) and patients with high expression, a rate of 88% (95% CI, 64–98; P =0.035). The univariate and multivariate analyses revealed no differences.

Overall survival. The Kaplan–Meier analysis revealed a trend when TAp73 levels were divided in quartiles because patients in the first lower quartiles had a 5-year OS rate of 70% (95% CI, 59–88) and patients in the 4th quartile, showing the higher TAp73 expression, a rate of 81% (95% CI, 65–89; P = 0.1). The univariate and multivariate analyses revealed no differences.

Ectopic expression of $\triangle Np73$ increases proliferation and drug resistance and modifies the levels of *HMGB1*, *ABCB1*, and *CASP1*

We transiently transfected HCT116 colon cancer cells with an expression vector containing $\Delta Np73$ or the empty vector. After 72 hours of transfection, a statistically significant increase in the cell number was observed in those cells ectopically expressing $\Delta Np73$ (Fig. 4A). The MTT cell proliferation assay also confirmed this fact (Fig. 4B). In addition, a significant difference in the initiation and rate of proliferation measured by the cell index and the slope of the curves in the RT-CES system was observed between

| Table 3. Univariate and multivariate analyses of the association between p73 isoforms and MDR1, HMG1, | |
|---|--|
| and caspase-1 expression and clinicopathologic characteristics and OS of colon cancer patients | |

| | Category | Univariate analysis | | | Multivariate analysis | | |
|---------------------------------------|-------------------|---------------------|------------|----------|-----------------------|------------|-------|
| Variable | | HR | 95% CI | Р | HR | 95% CI | Р |
| Stage | l vs. II | 1.6 | 0.19–12.86 | 0.66 | 0.57 | 0.06–5.5 | 0.63 |
| | l vs. III | 8.8 | 1.13–68.5 | 0.037 | 7.18 | 0.86-60.1 | 0.069 |
| | I vs. IV | 75.6 | 7.47-764.8 | < 0.0001 | 26.5 | 2.42-289.9 | 0.007 |
| Vascular invasion | Yes vs. no | 5.13 | 2.23-11.81 | < 0.0001 | | | |
| Tumor differentiation | Well vs. poor | 2.29 | 0.83-6.3 | 0.11 | | | |
| | Moderate vs. poor | 1.69 | 0.21-13.64 | 0.62 | | | |
| LNM | Yes vs. no | 7.23 | 3.19–16.37 | < 0.0001 | | | |
| Bicentiles ∆ <i>Ex2p73</i> expression | Low vs. high | 1.53 | 0.7–3.35 | 0.28 | | | |
| Bicentiles ∆Ex2/3p73 expression | Low vs. high | 2.28 | 1.02-5.1 | 0.044 | | | |
| Bicentiles $\Delta Np73$ expression | Low vs. high | 2.1 | 0.94-4.68 | 0.07 | | | |
| Bicentiles TAp73 expression | Low vs. high | 0.73 | 0.34-1.56 | 0.4 | | | |
| Bicentiles ABCB1 expression | Low vs. high | 2.28 | 1.04-4.99 | 0.04 | 4.5 | 1.48–13.92 | 0.008 |
| Bicentiles HMGB1 expression | High vs. low | 2.61 | 0.99–6.9 | 0.05 | 6.25 | 1.61–24.19 | 0.008 |
| Bicentiles CASP1 expression | Low vs. high | 0.70 | 0.31–1.57 | 0.39 | | | |

NOTE: The blank cells correspond to variables that showed no independent relationship to OS in the multivariate analysis. Abbreviation: LNM, lymph node metastasis.

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Figure 4. Ectopic expression of ANp73 increases the proliferation of HCT116 colon cancer cells 72 hours after transfection. A statistically significant increase in the number of cells. Experiments were done in guadruplicates and counted in a cell counter apparatus (Digital Bio). B, MTT assav shows that ectopic expression of $\Delta Np73$ leads to an increase in the cell-proliferation rate compared with the mock HCT116 cells (*, P < 0.001; *, P < 0.0001). C, significant difference in the initiation and rate of proliferation measured by the cell index and the scope of the curves in the RT-CES system (***, P < 0.0001. P value was calculated taking the different cell-index measurements in the exponential cellular growth phase).

both cells (Fig. 4C). The ectopic expression of Δ Np73 does not compromise the viability of the cells, being in both cell types, with cells overexpressing the isoform and the control variant in the range of 95% to 98%.

Cells expressing the $\Delta Np73$ vector showed 30% higher viability after oxaliplatin exposure than those transfected with the mock vector (Fig. 5). Oxaliplatin did not modify the endogenous levels of $\Delta Np73$.

No modification in migration and invasion was detected.

Ectopic expression of $\Delta Np73$ led to a 6- to 20-fold increase in its mRNA levels compared with the mock vector. This increase was accompanied by an upregulation in the mRNA levels of *ABCB1* and *HMGB1* of 2- to 8-fold. No modifications in CASP1 levels were detected.

Discussion

Although several studies have linked the upregulation of specific *TP73* isoforms with poor tumor prognosis parameters (20), little information is available on the impact of the altered expression of *TP73* variants on patient survival. $\Delta Np73$ overexpression is associated with shorter survival in patients with neuroblastoma (27), meduloblastoma (28), and lung (26), hepatocellular (30), and cervical squamous

cell carcinomas (29). In addition, $\Delta Ex2/3p73$ variant upregulation is associated with survival in patients with lowgrade glioma (31). In our colon cancer patient series, we observed that overexpression of $\Delta Ex2/3p73$ and $\Delta Np73$ forms predict OS, although only the pathologic stage remains an independent predictor in the multivariate analysis. Two reports have described the upregulation of TP73 as an independent marker of colorectal cancer patient survival (23, 52). These publications analyzed the general levels of TP73 without taking into consideration the different variants that could really be involved in the shortening of survival. Although the general levels of TP73 could be used in the clinical setting as a survival predictor, there has recently been increasing focus on unraveling which specific TP73 isoforms really support the oncogenic role in human cancer processes. Our results point to $\Delta Ex2/3p73$ and $\Delta Np73$ as the variants that may contain these oncogenic properties. Intriguingly, we have observed that those cases with concomitant overexpression of specific TP73 isoforms and inactive TP53 showed a better outcome. It may be possible that the inactivation of TP53 through mutation could trigger specific tumor-suppressor pathways that might partially compensate for the oncogenic environment generated by the overexpression of the Δ TAp73 variants. It is mandatory to unravel the complex mechan-

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isms involved in the simultaneous regulation of target genes by TP53, TAp73, and ΔNp73 isoforms and the putative feedback among them to obtain solid conclusions from the cancer patient studies. Lastly, it is interesting to note that both TAp73 and Δ TAp73 forms were found to be upregulated in a significant number of our colon tumors. It is possible that the presence of Δ TAp73 variants, even at low levels, completely suppresses the transactivation activity of TAp73, with the consequent elimination of essential TAp73 antitumorigenic function. Furthermore, at the protein level, $\Delta TAp73$ isoforms have been described to be more stable than those of TAp73, in terms of what can contribute to promote a cellular oncogenic context (2, 13). The use of compounds that can increase the stability of TAp73 variants, such as netrin-1 (53), could diminish this tumorigenic environment.

The association of the overexpression of $\Delta TAp73$ isoforms with shorter survival could be due to some putative TP73 target genes being involved in drug resistance, invasiveness, and other stages of the tumorigenesis process. ABCB1, HMGB1, and CASP1, among others, have been described as TP73 targets (35-38). These previous data are supported by the fact that the ectopic expression of $\Delta Np73$ in our cellular system induces the upregulation of ABCB1 and HMGB1. In our study, direct statistical correlation was found between expression of $\Delta Ex2p73$, $\Delta Ex2/3p73$, and $\Delta Np73$ and HMGB1 levels. Furthermore, a direct trend was observed between the same variants and ABCB1 expression levels. This supports the possible positive regulation of *HMGB1* and *ABCB1* by the $\Delta TAp73$ forms *in vivo* in colorectal carcinomas. In a larger colon cancer patient series, the correlation between ABCB1 and $\Delta TAp73$ variants might reach statistical significance. Although TP73 has been described as regulating CASP1 expression, no such direct correlation between CASP1 levels and TAp73 expression was found in our set of patients (38).

In addition, *ABCB1* and *HMGB1* overexpression was associated with shorter OS of patients. In the multivariate analysis including clinicopathologic parameters of the

tumors and the levels of TP73 variants, ABCB1, HMGB1, and CASP1, we observed that, in addition to tumor stage, ABCB1 and HMGB1 expression were also strong, independent predictors of OS. These data underline the importance of identifying the specific targets downstream of Δ TAp73 isoforms, which might have an oncogenic function and could be stronger than TP73 variants themselves in predicting patient outcome. As such, they could be used as prognostic markers in the clinical setting. Little is known about the relevance of ABCB1 and HMGB1 to the outcome of cancer patients, although the fact that upregulation of ABCB1 and HMGB1 has been associated in a few reports with poor prognosis of cancer patients sustains our hypothesis (44-46). Remarkably, the finding that the ectopic expression of $\Delta Np73$ increases the proliferation rate and confers resistance to oxaliplatin to colon cancer cells strengthens the oncogenic potential of this specific isoform and its involvement in specific tumorigenesis processes. As previously reported by other groups (54, 55), the exposure of the cells to oxaliplatin did not modify the endogenous levels of TP73 variants. It is possible that those tumors already expressing high levels of $\Delta Np73$ can show resistance to the treatment; in addition, the oxaliplatin action could gradually select the cells overexpressing this putative oncogenic p73 variant, resulting in a resistant tumor.

As cumulative data support the oncogenic role of Δ TAp73 isoforms (14, 56), the mechanisms and targets underlying these functions are currently of great interest. In this article, we present original data with regard to the impact of specific *TP73* variants in the outcome of colon cancer patients. In addition, we found that putative Δ TAp73 isoform targets are independent prognostic markers of OS. Specifically, upregulation of *ABCB1* and *HMGB1* predicts, in a strong, independent manner, the OS of patients diagnosed with colon cancer. Further experiments are needed to identify specific targets of Δ TAp73 isoforms that carry out an oncogenic role and could be used as clinical markers of relapse.

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Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed

Acknowledgments

We thank Mike Eaude for his assistance with correction of the language in the manuscript.

References

- Jost CA, Marin MC, Kaelin WG Jr. p73 is a simian [correction of human] p53-related protein that can induce apoptosis. Nature 1997;389:191–4.
- Stiewe T, Theseling CC, Putzer BM. Transactivation-deficient Delta TA-p73 inhibits p53 by direct competition for DNA binding: implications for tumorigenesis. J Biol Chem 2002;277:14177–85.
- Stiewe T, Stanelle J, Theseling CC, Pollmeier B, Beitzinger M, Pützer BM. Inactivation of retinoblastoma (Rb) tumor suppressor by oncogenic isoforms of the p53 family member p73. J Biol Chem 2003;278: 14230–6.
- Zaika AI, Slade N, Erster SH, Sansome C, Joseph TW, Pearl M, et al. ∆Np73, a dominant-negative inhibitor of wild-type p53 and TAp73, is up-regulated in human tumors. J Exp Med 2002;196:765–80.
- Pozniak CD, Radinovic S, Yang A, McKeon F, Kaplan DR, Miller FD. An anti-apoptotic role for the p53 family member, p73, during developmental neuron death. Science 2000;289:304–6.
- Stiewe T, Zimmermann S, Frilling A, Esche H, Pützer BM. Transactivation-deficient ∆TA-p73 acts as an oncogene. Cancer Res 2002;62: 3598–602.
- Petrenko O, Zaika A, Moll UM. ΔNp73 facilitates cell immortalization and cooperates with oncogenic ras in cellular transformation *in vivo*. Mol Cell Biol 2003;23:5540–55.
- Irwin MS, Kaelin WG. p53 family update: p73 and p63 develop their own identities. Cell Growth Differ 2001;12:337–49.
- Melino G, De Laurenzi V, Vousden KH. p73: friend or foe in tumorigenesis. Nat Rev Cancer 2002;2:605–15.
- Zhu J, Jiang J, Zhou W, Chen X. The potential tumor suppressor p73 differentially regulates cellular p53 target genes. Cancer Res 1998;58: 5061–5.
- Lin Y-L, Sengupta S, Gurdziel K, Bell GW, Jacks T, Flores ER. p63 and p73 transcriptionally regulate genes involved in DNA repair. PLoS Genet 2009;5:e1000680.
- Scian MJ, Carchman EH, Mohanraj L, Stagliano KE, Anderson MA, Deb D, et al. Wild-type p53 and p73 negatively regulates expression of proliferation related genes. Oncogene 2008;27:2583–93.
- Grob TJ, Novak U, Maisse C, Barcaroli D, Lüthi AU, Pirnia F. Human ∆Np73 regulates a dominant negative feedback loop for TAp73 and p53. Cell Death Differ 2001;8:1213–23.
- Wilhelm MT, Rufini A, Wetzel MK, Tsuchihara K, Inoue S, Tomasini R, et al. Isoform-specific p73 knockout mice reveal a novel role for DeltaNp73 in the DNA damage response pathway. Genes Dev 2010;24:549–60.
- Donehower LA, Harvey M, Slagle BL, McArthur MJ, Montgomery CA Jr, Butel JS, et al. Mice deficient for p53 are developmentally normal but susceptible to spontaneous tumours. Nature 1992; 356:215–21.
- Yang A, Walker N, Bronson R, Kaghad M, Oosterwegel M, Bonnin J, et al. p73-deficient mice have neurological, pheromonal and inflammatory defects but lack spontaneous tumours. Nature 2000;404: 99–103.
- Tomasini R, Tsuchihara K, Wilhelm M, Fujitani M, Rufini A, Cheung CC, et al. TAp73 knockout shows genomic instability with infertility and tumor suppressor functions. Genes Dev 2008;22:2677–91.
- Concin N, Becker K, Slade N, Erster S, Mueller-Holzner E, Ulmer H, et al. Transdominant ∆TAp73 isoforms are frequently up-regulated in ovarian cancer. Evidence for their role as epigenetic p53 inhibitors *in vivo*. Cancer Res 2004;64:2449–60.

Grant Support

This study was supported by FIS: PI08/0605; ISCIII-RD06/0020/0020; S-GEN-266-2006; Fundación Científica AECC; and SAF2007-60214.

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Received September 13, 2010; revised July 20, 2011; accepted July 20, 2011; published OnlineFirst August 1, 2011.

- Stiewe T, Tuve S, Peter M, Tannapfel A, Elmaagacli AH, Pützer BM. Quantitative TP73 transcript analysis in hepatocellular carcinomas. Clin Cancer Res 200410:626–33.
- 20. Domínguez G, García JM, Peña C, Silva J, García V, Martínez L, et al. ΔTAp73 upregulation correlates with poor prognosis in human tumors: Putative *in vivo* network involving p73 isoforms, p53 and E2F-1. J Clin Oncol 2006;24:805–15.
- Tuve S, Wagner SN, Schittek B, Pützer BM. Alterations of ∆TA–p73 splice transcripts during melanoma development and progression. Int J Cancer 2004;108:162–6.
- Leupin N, Luthi A, Novak U, Grob TJ, Hügli B, Graber H, et al. p73 status in B-cell chronic lymphocytic leukaemia. Leuk Lymphoma 2004;45:1205–7.
- Sun XF. p73 overexpression is a prognostic factor in patients with colorectal adenocarcinoma. Clin Cancer Res 2002;8:165–70.
- Becker K, Pancoska P, Concin N, Vanden Heuvel K, Slade N, Fischer M, et al. Patterns of p73 N-terminal isoform expression and p53 status have prognostic value in gynaecological cancers. Int J Oncol 2006;29:889–902.
- 25. Concin N, Hofstetter G, Berger A, Gehmacher A, Reimer D, Watrowski R, et al. Clinical relevance of dominant-negative p73 isoforms for responsiveness to chemotherapy and survival in ovarian cancer: evidence for a crucial p53-p73 cross-talk *in vivo*. Clin Cancer Res 2005;11:8372–83.
- Uramoto H, Sugio K, Oyama T, Nakata S, Ono K, Morita M, et al. Expression of deltaNp73 predicts prognosis in lung cancer. Clin Cancer Res 2004;10:6905–11.
- Casciano I, Mazzocco K, Boni L, Pagnan G, Banelli B, Allemanni G, et al. Expression of DeltaNp73 is a molecular marker for adverse outcome in neutoblastoma patients. Cell Death Differ 2002;9:246–51.
- 28. Zitterbart K, Zavrelova I, Kadlecova J, Spesna R, Kratochvilova A, Pavelka Z, et al. p73 expression in medulloblastoma: TAp73/DeltaNp73 transcript detection and possible association of p73alpha/ DeltaNp73 immunoreactivity with survival. Acta Neuropathol 2007; 114:641–50.
- 29. Liu SS, Chan KY, Cheung AN, Liao XY, Leung TW, Ngan HY. Expression of deltaNp73 and TAp73alpha independently associated with radiosensitivities and prognoses in cervical squamous cell carcinoma. Clin Cancer Res 2006;12:3922–7.
- Müller M, Schilling T, Sayan AE, Kairat A, Lorenz K, Schulze-Bergkamen H, et al. TAp73/DeltaNp73 influences apoptotic response, chemosensitivity and prognosis in hepatocellular carcinoma. Cell Death Differ 2005;12:1564–77.
- Wager M, Guilhot J, Blanc JL, Ferrand S, Milin S, Bataille B, et al. Prognostic value of increase in transcript levels of Tp73 DeltaEx2-3 isoforms in low-grade glioma patients. Br J Cancer 2006;95:1062–9.
- Lowe SW, Ruley HE, Jacks T, Housman DE. p53-dependent apoptosis modulates the cytotoxicity of anticancer agents. Cell 1993;74: 957–67.
- Irwin MS, Kondo K, Marin MC, Seelan RS, Smith DJ, Liu W, et al. Chemosensivity linked to p73 function. Cancer Cell 2003;3:403–10.
- Lunghi P, Costanzo A, Mazzera L, Rizzoli V, Levrero M, Bonati A. The p53 family protein p73 provides new insights into cancer chemosensitivity and targeting. Clin Cancer Res 2009;15:6495–502.
- 35. Vilgelm A, Wei JX, Piazuelo MB, Washington MK, Prassolov V, El-Rifai W, et al. DeltaNp73 alpha regulates MDR1 expression by inhibiting p53 function. Oncogene 2008;27:2170–6.

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- Johnson RA, Shepard EM, Scotto KW. Differential regulation of MDR1 transcription by the p53 family members. J Biol Chemistry 2005;280: 13213–9.
- 37. Uramoto H, Izumi H, Nagatani G, Ohmori H, Nagasue N, Ise T, et al. Physical interaction of tumor suppressor p53/p73 with CCAAT-binding transcription factor 2(CFT1) and differential regulation of human high-mobility group 1 (HMG1) gene expression. Biochem J 2003;371: 301–10.
- Jain N, Gupta S, Sudhakar C, Radha V, Swarup G. Role of p73 in regulating human caspase-1 gene transcription induced by interferongamma and cisplatin. J Biol Chem 2005;280:36664–73.
- Ambudkar SV, Kimchi-Sarfaty C, Sauna ZE, Gottesman MM. Pglycoprotein: from genomics to mechanism. Oncogene 2003;22: 7468–85.
- Tomas JO, Travers AA. HMG1 and 2, and related " architectural" DNAbinding proteins. Trends Biochem Sci 2001;26:167–74.
- Dengryse B, Bonaldi T, Scaffidi P, Müller S, Resnati M, Sanvito F, et al. The high mobility group (HMG) boxes of the nuclear protein HMG1 induce chemotaxis and cytoskeleton reorganization in rat smooth muscle cells. J Cell Biol 2001;152:1197–206.
- 42. Jarry A, Vallete G, Cassagnau E, Moreau A, Bou-Hanna C, Lemarre P, et al. Interleukin 1 and interleukin 1beta converting enzyme (caspase 1) expression in the human colonic epithelial barrier. Caspase 1 downregulation in colon cancer. Gut 1999;45:246–51.
- **43.** Winter RN, Kramer A, Borkowski A, Kyprianou N. Loss of capase-1 and caspase-3 protein expression in human prostate cancer. Cancer Res 2001;61:1227–32.
- 44. Sinicrope FA, Hart J, Brasitus TA, Michelassi F, Lee JJ, Safa AR. Relationship of P-glycoprotein and carcinoembryonic antigen expression in human colon carcinoma to local invasion, DNA ploidy, and disease relapse. Cancer 1994;74:2908–17.
- 45. Wu D, Ding Y, Wang S, Zhang Q, Liu L. Increased expression of high mobility group box 1 (HMGB1) is associated with progression and poor prognosis in human nasopharyngeal carcinoma. J Pathol 2008;216:167–75.
- 46. Yao X, Zhao G, Yang H, Hong X, Bie L, Liu G. Overexpression of highmobility group box 1 correlates with tumor progression and poor

prognosis in human colorectal carcinoma. J Cancer Res Clin Oncol 2010;136:677–84.

- 47. Abassi YA, Xi B, Zhang W, Ye P, Kirstein SL, Gaylord MR, et al. Kinetic cell-based morphological screening: prediction of mechanism of compound action and off-target effects. Chem Biol 2009;16:712–23.
- 48. Sjögren S, Inganäs M, Torbjörn N, Lindgren A, Nordgren H, Holmberg L, et al. The p53 gene in breast cancer: Prognostic value of complementary DNA sequencing versus immunohistochemistry. J Natl Cancer Inst 1996;88:173–82.
- Kaplan EL, Meier P. Non-parametric estimation from incomplete observations. J Am Stat Assoc 1958;53:457–81.
- Peto R, Pike MC, Armitage P, Breslow NE, Cox DR, Howard SV, et al. Design and analysis of randomized clinical trials requiring prolonged observations of each patient: II. Analysis and examples. Br J Cancer 1977;35:1–39.
- 51. Cox DR. Regression models and life tables. J Stat Soc 1972;34: 187-20.
- 52. Toumi AA, El Hadj Oel A, Ben Mahmoud LK, Ben Hmida Ael M, Chaar I, Gharbi L, et al. The prognostic value of p73 overexpression in colorectal carcinoma: a clinicopathologic, immunohistochemical, and statistical study of 204 patients. Appl Immunohistochem Mol Morphol 2010;18:128–36.
- Roperch JP, El Ouadrani K, Hendrix A, Emami S, De Wever O, Melino G, et al. Netrin-1 induces apoptosis in human cervical tumor cells via the TAp73alpha tumor suppressor. Cancer Res 2008;68:8231–9.
- 54. Koivusalo R, Krausz E, Ruotsalainen P, Helenius H, Hietanen S. Chemoradiation of cervical cancer cells: Targeting human papillomavirus E6 and p53 leads to either augmented or attenuated apoptosis depending on the platinum carrier ligand. Cancer Res 2002;62:7364–71.
- 55. Wakasugi T, Izumi H, Uchiumi T, Suzuki H, Arao T, Nishio K, et al. ZNF143 interacts with p73 and is involved in cisplatin resistance through the transcriptional regulation of DNA repair genes. Oncogene 2007;26:5194–203.
- 56. Schuster A, Schilling T, De Laurezi V, Koch AF, Seitz S, Staib F, et al. $\Delta Np73\beta$ is oncogenic in hepatocellular carcinoma by blocking apoptosis signaling via death receptors and mitochondria. Cell Cycle 2010;9:2629–39.

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